

Bone marrow stromal cell transplantation for ischemic stroke — its multi-functional feature

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In this article, the author reviews recent advancements of basic research on bone marrow stromal cell (BMSC) transplantation for ischemic stroke. The BMSCs are easily isolated from the patients themselves and transplanted into them without any ethical and immunological problem. Animal experiments have shown that BMSC transplantation significantly enhance the recovery of motor and/or cognitive function in various types of neurological disorders such as ischemic stroke. The transplanted BMSCs aggressively migrate toward the damaged tissue and proliferate in the host brain. The BMSCs significantly improve the neuronal receptor function and local glucose metabolism in the peri-infarct area when transplanted into the infarct brain. Recent studies strongly suggest that the BMSCs contain heterogeneous subpopulations and contribute to functional recovery through multiple mechanisms, including neuroprotection, inflammatory modulation, cell fusion, and neural differentiation. The author describes the importance to establish BMSC transplantation as a therapeutic entity that is scientifically proven.

Key words: bone marrow stromal cell, transplantation, cell therapy, ischemic stroke

INTRODUCTION

There are few drugs to effectively protect or repair the central nervous system (CNS) in clinical situation in spite of the huge efforts to develop them for longer than 50 years (Savitz and Fisher 2007). As the results, cell therapy has recently been expected as the alternative treatment strategy to enhance functional recovery after various kinds of neurological disorders, including ischemic stroke and spinal cord injury. Previously, a variety of cells have been studied as the candidates of donor cells for this purpose. These include embryonic stem (ES) cells, neural stem cells, induced pluripotent stem (iPS) cells, umbilical cord blood cells, and bone marrow stromal cells (BMSCs) (Jablonska and Lukomska 2011). Of these, the BMSCs may have the most enormous therapeutic potential among them, because they can be obtained from the patients themselves and easily expanded without posing any ethical and immunological prob-

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lems. The BMSCs are non-hematopoietic cells and are also known as mesenchymal stromal cells (MSCs). For the decades, numerous numbers of studies have indicated that the transplanted BMSCs significantly enhance functional recovery after the insults in animal models of various neurological disorders. For example, the BMSCs significantly enhance the recovery of motor function when transplanted into the animal models of cerebral infarct, SCI, and TBI (Bliss et al. 2007, Parr et al. 2007). More interestingly, the BMSCs have the potential to ameliorate cognitive dysfunction under certain conditions. Thus, Wu and coauthors (2007) directly transplanted the BMSC into the hippocampus and found significant improvement of cognitive function in Alzheimer' disease model of rats. Maruichi and others (2009) stereotactically transplanted the BMSC into the mice subjected to diffuse axonal injury, and concluded that BMSC transplantation significantly enhance the recovery of cognitive function on Morris Water Maze test. Furthermore, Shichinohe and colleagues (2010) have also demonstrated that the BMSC significantly ameliorate white matter damage and improve cognitive function in chronic cerebral ischemia model of rats.

Based on these preclinical results, some of preliminary clinical testing has already been conducted to evaluate the safety and therapeutic effects of BMSC transplantation for the patients with both acute and chronic neurological disorders (Bang et al. 2005, Lee et al. 2008, Zhang et al. 2008, Pal et al. 2009, Lee et al. 2010, Mazzini et al. 2010, Saito et al. 2012). However, it should be reminded that a variety of questions or problems still remains to be solved in order to establish BMSC transplantation as scientifically proven entity in clinical situation (Abe et al. 2012). This article reviews recent knowledge on basic aspects of BMSC transplantation for ischemic stroke.

Mechanisms of CNS protection and repair by BMSCs

Recent studies have shed light on the mechanisms through which the BMSCs enhance functional recovery after cerebral infarct. Thus, Shichinohe and coworkers (2006) reported that BMSC transplantation significantly improved the binding potential for 125I-iomazenil, a specific ligand for γ-aminobutyric acid (GABA) receptor, in the periinfarct area. Mori and colleagues (2005) also showed that the engrafted BMSCs also improve glucose metabolism in response to sensory stimuli when transplanted into the rat cold injury model. Very recent study has demonstrated that the BMSCs may enhance functional recovery by promoting the recovery of local glucose metabolism in the periinfarct area when directly transplanted into the infarct brain (Miyamoto et al. 2012).

Furthermore, biological or molecular roles of the BMSCs in the CNS have recently been elucidated. As first reported by Friedenstein and coauthors (1976), the BMSCs can be isolated using their biological properties to adhere to tissue culture surfaces. The adherent cells are well known to differentiate into osteoblast, chondrocytes, adipocytes, cardiomyocytes, and neural cells (Friedenstein et al. 1976, Prockop et al. 2003). However, they are morphologically heterogeneous. Therefore, it is quite natural to hypothesize that the BMSCs are the mixture of biologically various subpopulations of cells and contribute to enhance functional recovery through multiple mechanisms. In fact, our recent study has proven it (see below) (Hokari et al. 2008).

Migration and proliferation of BMSCs

The transplanted BMSC are known to aggressively migrate towards the lesion, although the underlying mechanisms are not clarified. Recent studies have shown that some chemokine such as monocyte chemoattractant protein-1 (MCP-1) and stromal cellderived factor (SDF)-1α are expressed around the damaged CNS tissue and play an important role in the migration of the transplanted cells (Wang et al. 2002, Askari et al. 2003). Recently, CXCR4, a specific receptor for SDF-1α, are believed to play an important role in their migration in the CNS (Shichinohe et al. 2007). Son and coworkers (2006) also reported that SDF-1/ CXCR4 and HGF/c-Met axes were involved in the recruitment of BMSC to the damaged tissue. It may be quite valuable to elucidate the temporal profile of these chemokines around damaged CNS tissue to determine the optimal timing of BMSC transplantation.

There are few studies whether the engrafted BMSCs retain their proliferative activity in the host brain or not. Therefore, we labeled the GFP-expressing BMSCs with a superparamagnetic iron oxide (SPIO) agent and transplanted into the ipsilateral striatum of the mice infarct brain. Fluorescence immunohistochemistry revealed that many of the GFP-positive cells were widely distributed in the peri-infarct area and partially expressed MAP2 and NeuN at 3 months after transplantation. However, only a small number of SPIOpositive cells could be detected on Turnbull blue staining. Surprisingly, the ratio of the SPIO- to GFP-positive cells was less than 3%. The results strongly suggested that the BMSCs actively proliferate, toward the lesion, and partially express the neuronal phenotype in the host brain during 3 months after transplantation (Yano et al. 2005).

Nursing effects of BMSCs

The BMSCs may produce some neuroprotective or neurotrophic factors and support the survival of the host neural cells (Zhong et al. 2003). This hypothesis is readily reasonable because the BMSC *per se* support the homing and proliferation of the hematopoietic cells in the bone marrow by producing a variety of cytokines such as stromal cell-derived factor- 1α (SDF- 1α) (Kortesidis et al. 2005). Indeed, the conditioned medium of BMSCs significantly promote neurite outgrowth from the dorsal root ganglion (Neuhuber

et al. 2005). Recent study has clearly shown that the BMSCs release soluble neuroprotective factors, including nerve growth factor (NGF), hepatocyte growth factor (HGF) and brain-derived neurotrophic factor (BDNF), and significantly ameliorate glutamate-induced damage of neurons (Hokari et al. 2008). Furthermore, the BMSC-conditioned medium activates phosphorylation of mitogen-activated protein kinase/extracellular signal-regulated protein kinase and/or phosphoinositide 3-kinase/serine/threonine kinase (PI3K/Akt) in primary culture of rat dorsal root ganglion (DRG) neurons (Gu et al. 2009). The BMSCs markedly promote the neurite extension from the neurons in the organotypic slice of the brain and spinal cord (Kamei et al. 2007, Shichinohe et al. 2008). Hofstetter and colleagues (2002) transplanted the BMSC into the injured cord and found that the engrafted BMSC were tightly associated with longitudinally arranged immature astrocytes and formed bundles bridging the epicenter of the injury. Very recently, He and coauthors (2011) have reported that the BMSC significantly increase the expression of bFGF, BDNF, and vascular endothelial growth factor (VEGF) in the ischemic brain. These findings strongly suggest that the BMSCs trigger endogenous signaling pathways of survival and repair in neurons by secreting soluble neurotrophic factors (Gornicka-Pawlak et al. 2011).

Very recent studies have demonstrated the alternative pathways through which the BMSC may protect the neurons. Thus, Scheibe and others (2012) investigated the mechanism through which the BMSCs protect the neurons against oxygen-glucose deprivation model in vitro. They found that the BMSCs released plasminogen activator inhibitor (PAI)-1 and significantly improved neuronal survival by increasing the phosphorylation of STAT3 and Akt in the neurons (Scheibe et al. 2012). Nowadays, the neurovascular units (NVUs) are known quite important to maintaining the homeostasis in the CNS. The NVUs consist of endothelial cells, astrocytes, and neurons. The BMSCs also protect the neurovascular integrity between basement membrane and astrocyte end-feet and ameliorate brain damage in stroke-prone spontaneous hypertensive rats (SHR-SP) (Ito et al. 2012). Alternatively, it is well known that the BMSCs release the angiogenic factors such as VEGF and contribute to increase the vessel density in the ischemic organs (Hoffmann et al. 2010).

Immunomodulatory effects of BMSCs

Both neutrophils and macrophages are well known to play an important role in the early inflammation after cerebral infarct (Barone and Feuerstein 1999). Indeed, their inflammatory response may be an essential process to clear cellular debris and initiate the healing pathways. Simultaneously, however, these inflammatory reactions may also give rise to cytotoxic damage to the surviving neurons, astrocytes, and endothelial cells in the peri-infarct area (Barone and Feuerstein 1999).

On the other hands, the BMSCs have currently been investigated as donor cells for novel cell therapy to prevent and to treat clinical disease associated with aberrant immune response. Preclinical studies strongly suggest that the BMSCs may protect against infectious challenge either by direct effects on the pathogen or through indirect effects on the host. In the host, the BMSCs may attenuate pro-inflammatory cytokine and chemokine induction, reduce pro-inflammatory cell migration into sites of injury and infection, and induce immunoregulatory soluble and cellular factors to preserve organ function (Auletta et al. 2012). Based on these observations, the BMSCs have been expected as immunomodulators in tissue repair, autoimmune disease, and graft versus host disease (GVHD) (Mundra et al. 2012).

Interestingly, large numbers of mature neutrophils are retained near the BMSCs in the bone marrow, suggesting that the BMSCs protect these neutrophil pools from apoptosis and also prevent their inappropriate activation and release of granules to prevent accidental damage to the bone marrow (Bianco and Gehron Robey 2000, Raffaghello et al. 2008). The BMSCs also reduce their migration and release of reactive oxygen species (ROS) (van den Akker et al. 2013). Likewise, the BMSCs trigger the macrophage to go towards the anti-inflammatory phenotype and also reduce their release of pro-inflammatory cytokines such as interleukin (IL)-1, IL-6, and interferon (TNF)-y, while markedly increase their antiinflammatory cytokines such as IL-10 (van den Akker et al. 2013). Therefore, the transplanted BMSCs may prevent excessive inflammatory response and prevent further tissue damage in the peri-infarct area.

Cell fusion of BMSCs

Several studies have demonstrated that the BMSCs fuse with the host cells when they are transplanted into various kinds of organs or are co-cultured with donor cells (Terada et al. 2002, Alvarez-Dolado et al. 2003, Spees et al. 2003, Vassilopoulos et al. 2003). Spees and coauthors (2003) co-cultured the BMSCs with heatshocked human small airway epithelial cells, and found that about 25% of them fused with epithelial cells (Spees et al. 2003). We have found similar results when the Yang and colleagues (2012) recently reported that the BMSCs fuse with the hydrogen peroxidetreated cardiomyocytes and significantly ameliorate their apoptosis in vitro. They also found that the BMSCs highly fused with the cardiomyocytes when injected to the mice subjected to myocardial infarction (Yang et al. 2012). However, the function of the resulting hybrid cells should be further investigated to explore their roles in tissue protection (Curril et al. 2010). Very recently, Islam and coworkers (2012) reported that the BMSCs transfer their mitochondria to

the pulmonary alveolar epithelia through gap junction channels. The mitochondrial transfer increased alveolar ATP concentration and protected them against acute lung injury (Islam et al. 2012).

Neural differentiation of BMSCs

The BMSC per se are believed to differentiate into neural cells in the host's brain. This theory is based on the findings that BMSC simulate neuronal morphology and express the proteins specific for neurons in vitro (Sanchez-Ramos et al. 2000, Woodbury et al. 2000) or in vivo (Azizi et al. 1998, Kopen et al. 1999). Although the hypothesis is quite attractive, there still remain several questions. Actually, several studies posed a question about their in vitro differentiation into neurons (Lu et al. 2004, Neuhuber et al. 2004). Recent studies have shown that the BMSCs can alter their

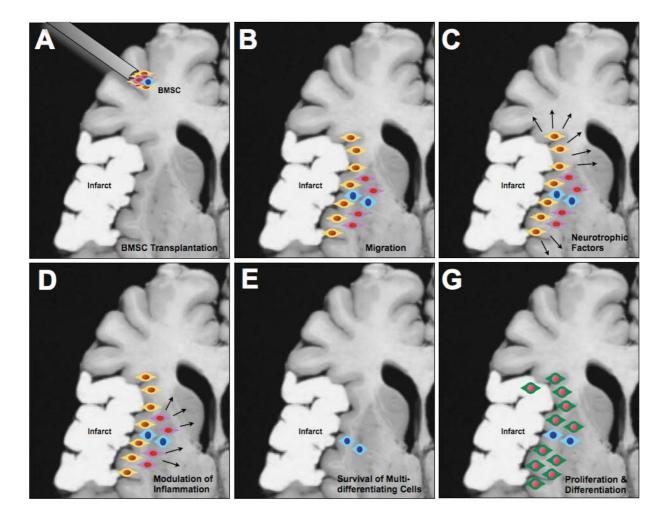


Fig. 1. Possible mechanism of functional recovery after ischemic stroke by bone marrow stromal cell (BMSC) transplantation

gene expression profile in response to exogenous stimuli and increase the genes related to the neural cells (Bossolasco et al. 2005, Hermann et al. 2006, Yamaguchi et al. 2006). Using microarray analysis, Yamaguchi and others (2006) showed that the BMSCs significantly reduce their genes related to mesenchymal cells and increased the neuron-related genes, when chemically treated with basic fibroblast growth factor (bFGF), retinoic acid (RA), and dimethyl sulfoxide (DMSO).

The BMSCs can acquire the neuronal phenotype under more physiological conditions. Thus, Sanchez-Ramos and colleagues (2000) showed that a small fraction of BMSCs cultured in epidermal growth factor (EGF) or retinoic acid/BDNF expressed nestin, NeuN, or GFAP, and that the proportion of NeuN-expressing cells increased when BMSC were co-cultured with fetal mouse midbrain neurons. Subsequently, Spees and others (2003) co-cultured the BMSCs with heatshocked small airway epithelial cells without any chemical agents, and found that BMSC rapidly differentiated into epithelial-like cells and repaired epithelial monolayer. Wislet-Gendebien and coauthors (2005) also co-cultured the BMSCs with cerebellar granule cells and assessed their fates. They found that the nestin-expressing BMSCs express other neuronal markers and that BMSC-derived neuron-like cells fire single-action potentials in response to neurotransmitters such as glutamate. Hokari and colleagues (2008) also demonstrated that a certain subpopulation of the BMSCs morphologically simulated the neuron and expressed the neuron-specific proteins without any evidence of cell fusion, when co-cultured with the neurons. These findings strongly suggest that at least a certain subpopulation of the BMSCs have the potential to alter their gene expression profile and to differentiate into the neural cells in response to the surrounding environment. In fact, the local environment may be the predominant determinant of the phenotypic fate of engrafted BMSCs in the host brain. Thus, the majority of them express the neuronal markers such as NeuN, MAP2, and Tuj-1 in the neocortex, while they express astrocytic phenotype in the corpus callosum or spinal cord (Lee et al. 2003, 2004, Shichinohe et al. 2007, Maruichi et al. 2009, Kawabori et al. 2012). The findings correlate very well with previous results. Shihabuddin and coworkers (2000) reported that adult spinal cord neural stem cells differentiated into neurons after transplantation into dentate gyrus of hip-

pocampus, but were unable to exhibit neurogenic potential when transplanted back into the adult spinal cord. Johansson and others (1999) also showed that neural progenitor cells start to proliferate, but differentiate into astrocytes after spinal cord injury. More importantly, the findings indicate that only the subgroup of BMSCs with potential of neural differentiation can survive in the host brain for a long time (>4 weeks).

More interestingly, recent study has shown that the engrafted BMSCs express γ-aminobutyric acid (GABA) receptor and improve the binding potential for 125Iiomazenil in the peri-infarct area (Shichinohe et al. 2006). Using micro-PET/CT apparatus, Miyamoto and colleagues (2013) serially quantified local glucose metabolism in the rat subjected to cerebral infarct and found that BMSC transplantation significantly enhance the recovery of glucose metabolism in the peri-infarct area. Alternatively, Chiba and coauthors (2009) have recently found that the BMSCs acquire neuronal phenotype and build synaptic connection with the corticospinal tract, when transplanted into the injured spinal cord of rats. *In vitro* studies have also indicated that the BMSCs exhibit electrical functions simulating those of neurons (Kohyama et al. 2001, Jiang et al. 2003, Jin et al. 2003), although this is still controversial (Hofstetter et al. 2002).

Very recently, Wakao and coworkers (2011) successfully isolated stress-tolerant adult human stem cells from cultured skin fibroblasts or BMSCs. These cells can self-renew, express a set of genes associated with pluripotency, and differentiate into endodermal, ectodermal, and mesodermal cells both in vitro and in vivo. When transplanted into immunodeficient mice by local or intravenous injection, they were integrated into damaged skin, muscle, or liver and differentiated into cytokeratin 14-, dystrophin-, or albumin-positive cells in the respective tissues. Furthermore, they can be efficiently isolated as SSEA-3-positive cells. Unlike authentic ES cells, their proliferation activity is not very high and they do not form teratomas in immunodeficient mouse testes. The findings are quite attractive, because non-tumorigenic stem cells with the ability to generate the multiple cell types of the three germ layers can be obtained through easily accessible adult human mesenchymal cells without introducing exogenous genes (Kuroda et al. 2010). These cells were named as multilineage-differentiating stress enduring (Muse) cells. Furthermore, they have proven that Muse

cells are a primary source of induced pluripotent stem (iPS) cells in human fibroblasts (Wakao et al. 2011). There results strongly suggest that a certain subpopulation of BMSCs may have the biological properties of neural differentiation and contribute to regenerate the infarct brain (Fig. 1).

CONCLUSION

Recent studies have gradually clarified the biological feature of BMSCs as the donor cells for ischemic stroke. The author emphasizes that it would be essential to fully explore it to apply BMSC transplantation into clinical situation.

REFERENCES

- Abe K, Yamashita T, Takizawa S, Kuroda S, Kinouchi H, Kawahara N (2012) Stem cell therapy for cerebral ischemia: from basic science to clinical applications. J Cereb Blood Flow Metab 32: 1317–1331.
- Alvarez-Dolado M, Pardal R, Garcia-Verdugo JM, Fike JR, Lee HO, Pfeffer K, Lois C, Morrison SJ, Alvarez-Buylla A (2003) Fusion of bone-marrow-derived cells with Purkinje neurons, cardiomyocytes and hepatocytes. Nature 425: 968–973.
- Askari AT, Unzek S, Popovic ZB, Goldman CK, Forudi F, Kiedrowski M, Rovner A, Ellis SG, Thomas JD, DiCorleto PE, Topol EJ, Penn MS (2003) Effect of stromal-cell-derived factor 1 on stem-cell homing and tissue regeneration in ischaemic cardiomyopathy. Lancet 362: 697–703.
- Auletta JJ, Deans RJ, Bartholomew AM (2012) Emerging roles for multipotent, bone marrow-derived stromal cells in host defense. Blood 119: 1801–1809.
- Azizi SA, Stokes D, Augelli BJ, DiGirolamo C, Prockop DJ (1998) Engraftment and migration of human bone marrow stromal cells implanted in the brains of albino ratssimilarities to astrocyte grafts. Proc Natl Acad Sci U S A 95: 3908–3913.
- Bang OY, Lee JS, Lee PH, Lee G (2005) Autologous mesenchymal stem cell transplantation in stroke patients. Ann Neurol 57: 874–882.
- Barone FC, Feuerstein GZ (1999) Inflammatory mediators and stroke: new opportunities for novel therapeutics. J Cereb Blood Flow Metab 19: 819–834.
- Bianco P, Gehron Robey P (2000) Marrow stromal stem cells. J Clin Invest 105: 1663–1668.
- Bliss T, Guzman R, Daadi M, Steinberg GK (2007) Cell transplantation therapy for stroke. Stroke 38: 817–826.

- Bossolasco P, Cova L, Calzarossa C, Rimoldi SG, Borsotti C, Deliliers GL, Silani V, Soligo D, Polli E (2005) Neuroglial differentiation of human bone marrow stem cells in vitro. Exp Neurol 193: 312–325.
- Chiba Y, Kuroda S, Maruichi K, Osanai T, Hokari M, Yano S, Shichinohe H, Hida K, Iwasaki Y (2009) Transplanted bone marrow stromal cells promote axonal regeneration and improve motor function in a rat spinal cord injury model. Neurosurgery 64: 991–999; discussion 999–1000.
- Curril IM, Koide M, Yang CH, Segal A, Wellman GC, Spees JL (2010) Incomplete reprogramming after fusion of human multipotent stromal cells and bronchial epithelial cells. FASEB J 24: 4856–4864.
- Friedenstein AJ, Gorskaja JF, Kulagina NN (1976) Fibroblast precursors in normal and irradiated mouse hematopoietic organs. Exp Hematol 4: 267–274.
- Gornicka-Pawlak E, Janowski M, Habich A, Jablonska A, Drela K, Kozlowska H, Lukomska B, Sypecka J, Domanska-Janik K (2011) Systemic treatment of focal brain injury in the rat by human umbilical cord blood cells being at different level of neural commitment. Acta Neurobiol Exp (Wars) 71: 46–64.
- Gu Y, Wang J, Ding F, Hu N, Wang Y, Gu X (2009) Neurotrophic actions of bone marrow stromal cells on primary culture of dorsal root ganglion tissues and neurons. J Mol Neurosci 40: 332–341
- He XY, Chen ZZ, Cai YQ, Xu G, Shang JH, Kou SB, Li M, Zhang HT, Duan CZ, Zhang SZ, Ke YQ, Zeng YJ, Xu RX, Jiang XD (2011) Expression of cytokines in rat brain with focal cerebral ischemia after grafting with bone marrow stromal cells and endothelial progenitor cells. Cytotherapy 13: 46–53.
- Hermann A, Liebau S, Gastl R, Fickert S, Habisch HJ, Fiedler J, Schwarz J, Brenner R, Storch A (2006) Comparative analysis of neuroectodermal differentiation capacity of human bone marrow stromal cells using various conversion protocols. J Neurosci Res 83: 1502–1514.
- Hoffmann J, Glassford AJ, Doyle TC, Robbins RC, Schrepfer S, Pelletier MP (2010) Angiogenic effects despite limited cell survival of bone marrow-derived mesenchymal stem cells under ischemia. Thorac Cardiovasc Surg 58: 136–142.
- Hofstetter CP, Schwarz EJ, Hess D, Widenfalk J, El Manira A, Prockop DJ, Olson L (2002) Marrow stromal cells form guiding strands in the injured spinal cord and promote recovery. Proc Natl Acad Sci U S A 99: 2199–2204.

- Hokari M, Kuroda S, Shichinohe H, Yano S, Hida K, Iwasaki Y (2008) Bone marrow stromal cells protect and repair damaged neurons through multiple mechanisms. J Neurosci Res 86: 1024-1035.
- Islam MN, Das SR, Emin MT, Wei M, Sun L, Westphalen K, Rowlands DJ, Quadri SK, Bhattacharya S, Bhattacharya J (2012) Mitochondrial transfer from bone-marrow-derived stromal cells to pulmonary alveoli protects against acute lung injury. Nat Med 18: 759-765.
- Ito M, Kuroda S, Sugiyama T, Maruichi K, Kawabori M, Nakayama N, Houkin K, Iwasaki Y (2012) Transplanted bone marrow stromal cells protect neurovascular units and ameliorate brain damage in stroke-prone spontaneously hypertensive rats. Neuropathology 32: 522–533.
- Jablonska A, Lukomska B (2011) Stroke induced brain changes: Implications for stem cell transplantation. Acta Neurobiol Exp (Wars) 71: 74–85.
- Jiang Y, Henderson D, Blackstad M, Chen A, Miller RF, Verfaillie CM (2003) Neuroectodermal differentiation from mouse multipotent adult progenitor cells. Proc Natl Acad Sci U S A 100 (Suppl 1): 11854-11860.
- Jin K, Mao XO, Batteur S, Sun Y, Greenberg DA (2003) Induction of neuronal markers in bone marrow cells: differential effects of growth factors and patterns of intracellular expression. Exp Neurol 184: 78-89.
- Johansson CB, Momma S, Clarke DL, Risling M, Lendahl U, Frisen J (1999) Identification of a neural stem cell in the adult mammalian central nervous system. Cell 96: 25-34.
- Kamei N, Tanaka N, Oishi Y, Ishikawa M, Hamasaki T, Nishida K, Nakanishi K, Sakai N, Ochi M (2007) Bone marrow stromal cells promoting corticospinal axon growth through the release of humoral factors in organotypic cocultures in neonatal rats. J Neurosurg Spine 6: 412-419.
- Kawabori M, Kuroda S, Sugiyama T, Ito M, Shichinohe H, Houkin K, Kuge Y, Tamaki N (2012) Intracerebral, but not intravenous, transplantation of bone marrow stromal cells enhances functional recovery in rat cerebral infarct: An optical imaging study. Neuropathology 32: 217–226.
- Kohyama J, Abe H, Shimazaki T, Koizumi A, Nakashima K, Gojo S, Taga T, Okano H, Hata J, Umezawa A (2001) Brain from bone: efficient "meta-differentiation" of marrow stroma-derived mature osteoblasts to neurons with Noggin or a demethylating agent. Differentiation 68: 235-244.
- Kopen GC, Prockop DJ, Phinney DG (1999) Marrow stromal cells migrate throughout forebrain and cerebellum, and they differentiate into astrocytes after injection into neo-

- natal mouse brains. Proc Natl Acad Sci U S A 96: 10711-10716.
- Kortesidis A, Zannettino A, Isenmann S, Shi S, Lapidot T, Gronthos S (2005) Stromal-derived factor-1 promotes the growth, survival, and development of human bone marrow stromal stem cells. Blood 105: 3793-3801.
- Kuroda Y, Kitada M, Wakao S, Nishikawa K, Tanimura Y, Makinoshima H, Goda M, Akashi H, Inutsuka A, Niwa A, Shigemoto T, Nabeshima Y, Nakahata T, Fujiyoshi Y, Dezawa M (2010) Unique multipotent cells in adult human mesenchymal cell populations. Proc Natl Acad Sci USA 107: 8639-8643.
- Lee JB, Kuroda S, Shichinohe H, Ikeda J, Seki T, Hida K, Tada M, Sawada K, Iwasaki Y (2003) Migration and differentiation of nuclear fluorescence-labeled bone marrow stromal cells after transplantation into cerebral infarct and spinal cord injury in mice. Neuropathology 23: 169–180.
- Lee JB, Kuroda S, Shichinohe H, Yano S, Kobayashi H, Hida K, Iwasaki Y (2004) A pre-clinical assessment model of rat autogeneic bone marrow stromal cell transplantation into the central nervous system. Brain Res Brain Res Protoc 14: 37-44.
- Lee JS, Hong JM, Moon GJ, Lee PH, Ahn YH, Bang OY (2010) A long-term follow-up study of intravenous autologous mesenchymal stem cell transplantation in patients with ischemic stroke. Stem Cells 28: 1099-1106.
- Lee PH, Kim JW, Bang OY, Ahn YH, Joo IS, Huh K (2008) Autologous mesenchymal stem cell therapy delays the progression of neurological deficits in patients with multiple system atrophy. Clin Pharmacol Ther 83: 723–730.
- Lu P, Blesch A, Tuszynski MH (2004) Induction of bone marrow stromal cells to neurons: differentiation, transdifferentiation, or artifact? J Neurosci Res 77: 174-191.
- Maruichi K, Kuroda S, Chiba Y, Hokari M, Shichinohe H, Hida K, Iwasaki Y (2009) Transplanted bone marrow stromal cells improves cognitive dysfunction due to diffuse axonal injury in rats. Neuropathology 29: 422–432.
- Mazzini L, Ferrero I, Luparello V, Rustichelli D, Gunetti M, Mareschi K, Testa L, Stecco A, Tarletti R, Miglioretti M, Fava E, Nasuelli N, Cisari C, Massara M, Vercelli R, Oggioni GD, Carriero A, Cantello R, Monaco F, Fagioli F (2010) Mesenchymal stem cell transplantation in amyotrophic lateral sclerosis: A Phase I clinical trial. Exp Neurol 223: 229-237.
- Miyamoto M, Kuroda S, Zhao S, Magota K, Shichinohe H, Houkin K, Kuge Y, Tamaki N (2012) Bone marrow stromal cell transplantation enhances recovery of local glucose metabolism after cerebral infarction in rats: A serial 18F-FDG PET study. J Nucl Med 54:145-150

- Mori K, Iwata J, Miyazaki M, Nakao Y, Maeda M (2005) Functional recovery of neuronal activity in rat whiskerbarrel cortex sensory pathway from freezing injury after transplantation of adult bone marrow stromal cells. J Cereb Blood Flow Metab 25: 887–898.
- Mundra V, Gerling IC, Mahato RI (2012) Mesenchymal stem cell-based therapy. Mol Pharm 10: 77–89.
- Neuhuber B, Gallo G, Howard L, Kostura L, Mackay A, Fischer I (2004) Reevaluation of in vitro differentiation protocols for bone marrow stromal cells: disruption of actin cytoskeleton induces rapid morphological changes and mimics neuronal phenotype. J Neurosci Res 77: 192–204.
- Neuhuber B, Timothy Himes B, Shumsky JS, Gallo G, Fischer I (2005) Axon growth and recovery of function supported by human bone marrow stromal cells in the injured spinal cord exhibit donor variations. Brain Res 1035: 73–85.
- Pal R, Venkataramana NK, Bansal A, Balaraju S, Jan M, Chandra R, Dixit A, Rauthan A, Murgod U, Totey S (2009) Ex vivo-expanded autologous bone marrow-derived mesenchymal stromal cells in human spinal cord injury/paraplegia: a pilot clinical study. Cytotherapy 11: 897–911.
- Parr AM, Tator CH, Keating A (2007) Bone marrow-derived mesenchymal stromal cells for the repair of central nervous system injury. Bone Marrow Transplant 40: 609–619.
- Prockop DJ, Gregory CA, Spees JL (2003) One strategy for cell and gene therapy: harnessing the power of adult stem cells to repair tissues. Proc Natl Acad Sci U S A 100 (Suppl 1): 11917–11923.
- Raffaghello L, Bianchi G, Bertolotto M, Montecucco F, Busca A, Dallegri F, Ottonello L, Pistoia V (2008) Human mesenchymal stem cells inhibit neutrophil apoptosis: a model for neutrophil preservation in the bone marrow niche. Stem cells 26: 151–162.
- Saito F, Nakatani T, Iwase M, Maeda Y, Murao Y, Suzuki Y, Fukushima M, Ide C (2012) Administration of cultured autologous bone marrow stromal cells into cerebrospinal fluid in spinal injury patients: a pilot study. Restor Neurol Neurosci 30: 127–136.
- Sanchez-Ramos J, Song S, Cardozo-Pelaez F, Hazzi C, Stedeford T, Willing A, Freeman TB, Saporta S, Janssen W, Patel N, Cooper DR, Sanberg PR (2000) Adult bone marrow stromal cells differentiate into neural cells in vitro. Exp Neurol 164: 247–256.
- Savitz SI, Fisher M (2007) Future of neuroprotection for acute stroke: in the aftermath of the SAINT trials. Ann Neurol 61: 396–402.

- Scheibe F, Klein O, Klose J, Priller J (2012) Mesenchymal stromal cells rescue cortical neurons from apoptotic cell death in an in vitro model of cerebral ischemia. Cellular and molecular neurobiology 32: 567–576.
- Shichinohe H, Kuroda S, Yano S, Ohnishi T, Tamagami H, Hida K, Iwasaki Y (2006) Improved expression of gamma-aminobutyric acid receptor in mice with cerebral infarct and transplanted bone marrow stromal cells: an autoradiographic and histologic analysis. J Nucl Med 47: 486–491.
- Shichinohe H, Kuroda S, Yano S, Hida K, Iwasaki Y (2007) Role of SDF-1/CXCR4 system in survival and migration of bone marrow stromal cells after transplantation into mice cerebral infarct. Brain Res 1183: 138–147.
- Shichinohe H, Kuroda S, Tsuji S, Yamaguchi S, Yano S, Lee JB, Kobayashi H, Kikuchi S, Hida K, Iwasaki Y (2008) Bone marrow stromal cells promote neurite extension in organotypic spinal cord slice: significance for cell transplantation therapy. Neurorehabil Neural Repair 22: 447–457.
- Shichinohe H, Kuroda S, Sugiyama T, Ito M, Kawabori M (2010) Bone marrow stromal cell transplantation attenuates cognitive dysfunction due to chronic cerebral ischemia in rats. Dement Geriatr Cogn Disord 30: 293–301.
- Shihabuddin LS, Horner PJ, Ray J, Gage FH (2000) Adult spinal cord stem cells generate neurons after transplantation in the adult dentate gyrus. J Neurosci 20: 8727–8735.
- Son BR, Marquez-Curtis LA, Kucia M, Wysoczynski M, Turner AR, Ratajczak J, Ratajczak MZ, Janowska-Wieczorek A (2006) Migration of bone marrow and cord blood mesenchymal stem cells in vitro is regulated by stromal-derived factor-1-CXCR4 and hepatocyte growth factor-c-met axes and involves matrix metalloproteinases. Stem Cells 24: 1254–1264.
- Spees JL, Olson SD, Ylostalo J, Lynch PJ, Smith J, Perry A, Peister A, Wang MY, Prockop DJ (2003) Differentiation, cell fusion, and nuclear fusion during ex vivo repair of epithelium by human adult stem cells from bone marrow stroma. Proc Natl Acad Sci U S A 100: 2397–2402.
- Terada N, Hamazaki T, Oka M, Hoki M, Mastalerz DM, Nakano Y, Meyer EM, Morel L, Petersen BE, Scott EW (2002) Bone marrow cells adopt the phenotype of other cells by spontaneous cell fusion. Nature 416: 542–545.
- van den Akker F, Deddens JC, Doevendans PA, Sluijter JP (2013) Cardiac stem cell therapy to modulate inflammation upon myocardial infarction. Biochim Biophys Acta 1830: 2449–2458.
- Vassilopoulos G, Wang PR, Russell DW (2003) Transplanted bone marrow regenerates liver by cell fusion. Nature 422: 901–904.

- Wakao S, Kitada M, Kuroda Y, Shigemoto T, Matsuse D, Akashi H, Tanimura Y, Tsuchiyama K, Kikuchi T, Goda M, Nakahata T, Fujiyoshi Y, Dezawa M (2011) Multilineage-differentiating stress-enduring (Muse) cells are a primary source of induced pluripotent stem cells in human fibroblasts. Proc Natl Acad Sci U S A 108: 9875-9880.
- Wang L, Li Y, Chen J, Gautam SC, Zhang Z, Lu M, Chopp M (2002) Ischemic cerebral tissue and MCP-1 enhance rat bone marrow stromal cell migration in interface culture. Exp Hematol 30: 831-836.
- Wislet-Gendebien S, Hans G, Leprince P, Rigo JM, Moonen G, Rogister B (2005) Plasticity of cultured mesenchymal stem cells: switch from nestin-positive to excitable neuron-like phenotype. Stem Cells 23: 392–402.
- Woodbury D, Schwarz EJ, Prockop DJ, Black IB (2000) Adult rat and human bone marrow stromal cells differentiate into neurons. J Neurosci Res 61: 364-370.
- Wu QY, Li J, Feng ZT, Wang TH (2007) Bone marrow stromal cells of transgenic mice can improve the cognitive ability of an Alzheimer's disease rat model. Neurosci Lett 417: 281-285.

- Yamaguchi S, Kuroda S, Kobayashi H, Shichinohe H, Yano S, Hida K, Shinpo K, Kikuchi S, Iwasaki Y (2006) The effects of neuronal induction on gene expression profile in bone marrow stromal cells (BMSC)-a preliminary study using microarray analysis. Brain Res 1087: 15-27.
- Yang WJ, Li SH, Weisel RD, Liu SM, Li RK (2012) Cell fusion contributes to the rescue of apoptotic cardiomyocytes by bone marrow cells. J Cell Mol Med 16: 3085-3095.
- Yano S, Kuroda S, Shichinohe H, Hida K, Iwasaki Y (2005) Do bone marrow stromal cells proliferate after transplantation into mice cerebral infarct? - a double labeling study. Brain Res 1065: 60-67.
- Zhang ZX, Guan LX, Zhang K, Zhang Q, Dai LJ (2008) A combined procedure to deliver autologous mesenchymal stromal cells to patients with traumatic brain injury. Cytotherapy 10: 134-139.
- Zhong C, Qin Z, Zhong CJ, Wang Y, Shen XY (2003) Neuroprotective effects of bone marrow stromal cells on rat organotypic hippocampal slice culture model of cerebral ischemia. Neurosci Lett 342: 93-96.