

## Alterations of GABA<sub>B</sub> binding caused by acute and chronic lead administration

## Jolanta Waśkiewicz

Department of Neurochemistry, Medical Research Centre, Polish Academy of Sciences, 3 Dworkowa St.,00-784 Warsaw, Poland



Abstract. The effect of lead on GABAB binding was studied on membranes obtained from acute and chronically lead-treated rats. Acute lead treatment reduced both GABAB affinity ( $K_D$ ) by about 30% and density of receptor ( $B_{max}$ ) by about 15%. On the contrary chronic lead treatment increased receptor capacity by about 20% in spite of decreased receptor affinity by 25%. Both acute and chronic lead treatment shiffted displacement curves toward higher concentration of non-labelled compound(decreased affinity). The results show that lead can affect the GABAB binding in two ways: by reducing affinity of binding and by altering capacity of binding.

**Key words:** GABA , gamma-aminobutyric acid,  $K_D$ , affinity of receptor,  $B_{\text{max}}$  , density of receptor

Lead ions are known to be toxic to the nervous system of man and animal (Davis et al. 1993). Both acute and chronic exposures to lead result in behavioural and neurological symptomes. In lead neurotoxicology an important issue relates to how lead interferes with chemical neurotransmission. Numerous investigations have been undertaken in order to explain the possible molecular mechanism of lead toxicity (Silbergeld 1982, Tschudy and Lamon 1990, Zareba and Chmielnicka 1992, Guilarte 1993, Jabońska et al. 1994). It has been postulated that Pb/Ca interactions may play an important role in the process (Silbergeld et al. 1974, Kolton and Yassi 1982).

Gamma-aminobutyric acid (GABA) is known as one of inhibitory neurotransmitters in the mammalian brain that induces the hyperpolarization of membrane potential by activating GABA receptor coupled with chloride ion channel (Olsen 1981). The observation that some effects of GABA in the CNS are not antagonized by the GABA receptor antagonist bicuculline (Hill and Bowery 1981), has led to the discovery of the GABAB class of GABA receptor, which is insensitive to bicuculline but sensitive to (-)-baclofen(Hill and Bowery 1981). It has been reported that the activation of GABAB receptor causes reduction of evoked transmitter release (Bowery et al. 1984).

Activation of GABA<sub>B</sub> receptors by baclofen or GABA increases membrane K<sup>+</sup> conductance post-synaptically and decreases Ca<sup>2+</sup> conductance presynaptically to depress transmitter release; these receptors are directly coupled to the K<sup>+</sup> and Ca<sup>2+</sup> channels through Gi/Go proteins and are modulated by guaninenucleotides (Bowery 1993).

The aim of this work was to assess the sensitivity of Ca-dependent GABA binding to lead toxicity, especially its influence on affinity and capacity of GABAB binding. As shown previously, Pb<sup>2+</sup> level in synaptosomal fraction (obtained from P<sub>2</sub> fraction) was more than twice higher than in control fraction (Strużyńska and Rafałowska 1994). Membranes for GABAB binding were obtained from crude P<sub>2</sub> fraction.

The study was performed on male Wistar rats. Two models of poisoning were used:

- 1. In the acute model 15 mg Pb(CH<sub>3</sub>COO)<sub>2</sub>/kg b.w. was injected intraperitoneally for 7 days into rats weighting 150 g. The control animals were injected with distilled water. The animals were decapitated and brains were removed.
- 2. In the chronic model of toxicity, lead acetate (200 mg/l H<sub>2</sub>O) was given to 3-week old rats in drinking water for 3 months. Control animals received distilled water without Pb(CH<sub>3</sub>COO)<sub>2</sub>. The mechanism of the gamma-aminobutyric acid receptor (GABA<sub>B</sub>) binding was examined using crude membrane fraction (P<sub>2</sub>) obtained from the rat brain according to Zukin et al. (1974). For the assay pellets (P<sub>2</sub> fraction) which had been frozen for at least 16 h at 15° C were allowed to thow for 20 min at room temperature before resuspension in Tris-HCl buffer (50 mM pH 7,4) plus CaCl<sub>2</sub> (2.5 mM).

The pellet obtained from the equivalent of one rat brain was resuspended in 10 ml Tris-HCl. The suspension was incubated for 45 min at 20°C before centrifugation at 7,000 g for 10 min. This working procedure was repeated three more times allowing 15 min incubation with addition of Tris-HCl plus CaCl<sub>2</sub> each time. The final pellet was resuspended in Tris-HCl buffer+CaCl<sub>2</sub> (~1 mg protein per 0.8 ml buffer plus 40 µM isoguvacine for the assay. Protein concentrations were determined by the method of Lowry et al. (1951). To each 0.8 ml aliquot of membrane suspension 0.1 ml Tris-HCl plus CaCl<sub>2</sub> containing various concentrations of unlabelled GABA and 0.1 ml Tris-HCl containing a fixed concentration(1-3 nM) of [<sup>3</sup>H]-GABA was added. The range of final concentrations of unlabelled GABA was 1 nM-100 µM. Nonspecific binding was determined by adding 100 µM baclofen. The mixture was incubated for 15 min at 20°C. Each incubation was terminated by filtration under vacuum through Whatman GF/B glass filter and then the filter was rinsed twice with 5 ml of ice-cold distilled water according to Ohmori et al. (1990). The radioactivity trapped on each filter was measured by a liquid scintillation spectrometer at a counting efficiency of 40%.

Acute lead treatment reduced  $GABA_B$  receptor affinity ( $K_D$ ) by about 30% and density of receptor ( $B_{max}$ ) by about 15% (Table I).

Kinetic parameters for [3H] GABA binding to GABAB receptors after acute lead treatment

Group	K <sub>D</sub> (nM)	B <sub>max</sub> (fmol/mg protein)
control lead treated	105±11 136±14	952±93 707±79

The values represent mean  $\pm$  SEM from 4 experiments; P < 0.05 (Student's t test).

Chronic lead treatment decreased receptor affinity (K<sub>D</sub>) by about 25% but increases receptor capacity (B<sub>max</sub>) by about 20% (Table II).

There was no significant difference between receptor affinity in acute and chronic lead treated animals. Decreased receptor affinity in both models of intoxication is illustrated by ligand displacement curves (Fig. 1. A and B), which they shiffted toward higher concentrations of non-labelled compound in both acute and chronic lead-treated material (decreased receptor affinity K<sub>D</sub>).

The mechanism through which lead affects behaviour and neurochemistry is unknown. It has been hypothesized that Pb<sup>2+</sup> and Ca<sup>2+</sup> may compe-

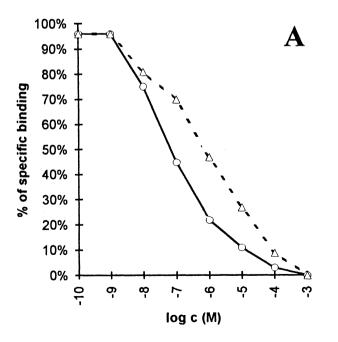
TABLE II

Kinetic parameters for [3H] GABA binding to the GABAB receptors after chronic lead treatment

Group	K <sub>D</sub> (nM)	B <sub>max</sub> (fmol/mg protein)
control	99±11	984±91
lead treated	126±12	1210±111

The results represent mean  $\pm$  SEM from 4 experiments; P < 0.05 (Student's t test).

titively interact at the sites involved in binding of GABA. Two hypotheses are proposed to explain these interactions: (1) competition with [Ca<sup>2+</sup>] in membrane Ca channels, which would inhibit the depolarization- induced influx of [Ca<sup>2+</sup>]; and/or (2) on intracellular [Ca<sup>2+</sup>] rise caused by inhibition of mitochondrial [Ca<sup>2+</sup>] fluxes (Silbergeld 1983). Our investigation on membrane fractions obtained from acute and chronic lead treatment rats confirm the latter hypothesis. Pb<sup>2+</sup> treatment of rats was associated with decrease affinity of GABA, binding and changes of GABA receptor density. As previously shown, chronic treatment of rat results in the inhibition of KCl dependent GABA release (Strużyńska



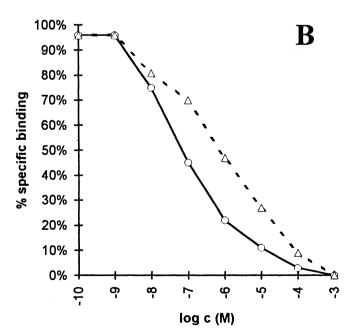


Fig. 1. Inhibition of 3 H-GABA binding by non-labelled GABA after (A)-acute and (B)-chronic treatment of lead . Control-(circles); Pb-treted-(triangles). The results are means of five experiments; the SEM was less than 5%.

and Rafałowska 1994). Our results are in agreement with these findings; lead inhibits GABA release and GABA affinity.

Decreased GABA receptor density in calcium dependent GABA binding in acute lead treated animals could be interpreted to manifest a decrease in number of the synapses and would be consistent with histological observations of brain tissue from lead treated animals (Krigman et al. 1974).

The variability in binding to lead treated membranes (changed receptor density) may result from the relative immaturity of this structure. Changes in chronic lead treated animals might be the consequence of changed protein synthesis which has been reported in both chronic and acute (Kennedy et al. 1983) lead studies and is consistent with the suggestion that a reduction in synapse number could result from a direct effect of lead on protein synthesis in chronically treated animals. Depression of protein synthesis and/or effects on lipid production (Krigman et al. 1974) by lead may also explain the decreased receptor affinity shown in acute and chronic lead treatment at calcium-dependent binding, which could result from increased production of endogennous receptor inhibitor rather than structural changes (Johnston 1981). The fact that different effects are observed in different types of neurones and at different times may reflect local differences in the concentration of free lead cations at critical developmental periods resulting from varying local pH and phosphate concentrations (Spence et al. 1985) in the different regions of brain. The solubility of lead complexes is dependent on pH and increases as pH declines (Bradbury 1979).

These investigations were supported by a Statutable grant from the State Committee for Scientific Research to Institute - Medical Research Centre.

- Bowery N.G.(1993) GABAB receptor pharmacology. Annu. Rev. Pharmacol.Toxicol. 33: 109-147.
- Bowery N.G., Price G.W., Hudson A.L., Hill D.R., Wilkin G.P., Turnbull M.J.(1984) GABA receptor multiplicity. Neuropharmacology 23: 219-231.
- Bradbury M.(1979) The concept of a blood-brain barrier.Wiley-Interscience, New York, p. 60-83.

- Davis J.M., Elias R.W., Grant L.D. (1993) Current issues in human lead exposure and regulation of lead. Neuro Toxicology 14: 15-28.
- Govoni S., Memo M., Lucchi L., Spano P.F. Trabucchi M. (1980) Brain neurotransmitter system and chronic lead intoxication. Pharm. Res. Commun. 12: 447-460
- Guilarte T.R.(1993) Neurochemical aspects of hippocampal and cortical lead neurotoxicity. Abstracts of 4th Meeting of the International Neurotoxicology, Elsinase, Denmark, p.12
- Hill D.R., Bowery N.G. (1981) <sup>3</sup>H-baclofen and <sup>3</sup>H-GABA bind to bicuculline-insensitive GABA<sub>B</sub> sites in rat brain. Nature 290: 149-152
- Jabłońska L., Walski M., Rafałowska U. (1994) Lead as an inductor of some morphological and functional changes in synaptosomes from rat brain. Cell. Mol. Neurobiol. 14: 701-709
- Johnston G.A.R.(1981) GABA receptors In:The role of peptides and amino acids as neurotransmitters (Eds. J.B. Lombardini and A.D. Kennny). Vol.68. Prog. Clin. Biol. Res. Liss, New York, p. 1-17.
- Karbon E.W., Duman R., Enna S.J.(1983) Biochemical identification of multiple GABA<sub>B</sub> binding sites:assocciation with noradrenergic terminals in rat forebrain. Brain Res. 274: 393-396
- Kennedy J.L., Girgis G.R., Rakhra S., McEwen Nicholls D. (1983) Protein synthesis in rat brain following neonatal exposure to lead. J. Neurol Sci. 59: 57-6 8
- Krigman M.R., Hogan E.L.(1974) Effect of lead intoxication on the postnatal growth of the rat nervous system. Envir. Hlth. Exp. Issue 7:187-189
- Lowry O.H., Rosebrough N.J., Farr A.L., Randall R.J.(1951)
  Protein measurement with the Folin phenol reagent. J.
  Biol. Chem. 1193: 265-275
- Ohmori Y., Hirouchi M., Taguchi J., Kuriyama K.(1990) Functional coupling of the gamma-aminobutyric acidB receptor with calcium ion channel and GTP-binding protein and its alteration of the gamma-aminobutyric acidB receptor. J. Neurochem. 54: 80-85
- Olsen R.W.(1981) GABA-benzodiazpine-barbiturate receptor interactions. J. Neurochem . 37: 1-13
- Silbergeld E.K. (1982) In Mechanisms of action of neurotoxic substances (Eds. K.N. Prasad and A. Vernadakis). Raven Press, New York, p. 1-25.
- Silbergeld E.K., Fales J.T., Goldberg A.M. (1974) The effects of inorganic lead on neuromuscular junction. Neuropharmacology 13: 795-801
- Spence I., Drew C., Johnston G., Lodge D. (1985) Acute effects of lead at central synapses in vitro. Brain Res.333: 103-109.
- Strużyńska L., Rafałowska U.(1994) The effect of lead on dopamine GABA and histidine spontaneous and KCl-dependent releases from rat brain synaptosomes. Acta Neurobiol. Exp. 54: 201-207

- Tschudy D.P., Lamon I.M. (1980) In Metabolic disease and control (8th ed.) (Eds. P.K. Bandy and L.E. Rosenberg). W.B. Saunders, Philadelphia, PA, p. 939-1007.
- Zareba G., Chmielnicka J. (1992) Disturbances in heme biosynthesis in rabbits after administration per os of low doses of tin and lead. Biol. Trace Element Res. 34: 115-122
- Zukin S.R., Young A..B., Snyder S.H. (1974) Gamma-aminobutyric acid binding to receptor sites in the rat central nervous system. Proc. Natl Acad. Sci. USA 71: 4802-4807

Paper presented at the 2nd International Congress of the Polish Neuroscience Society; Session: Neurochemistry